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Thiamine and its phosphate esters in relation to cardiometabolic risk factors in Saudi Arabs

Nasser M Al-Daghri^{1,2,3*}, Omar S Al-Attas^{1,2,3}, Khalid M Alkharfy^{1,2,4}, Majed S Alokail^{1,2}, Sherif H Abd-Alrahman¹ and Shaun Sabico¹

Abstract

Background: Thiamine deficiency has suggested to be linked to several insulin-resistance complications. In this study, we aim to associate circulating thiamine levels among cardiometabolic parameters in an Arab cohort using a simple, sensitive, rapid and selective high-performance liquid chromatography (HPLC) method that has recently been developed.

Methods: A total of 236 randomly selected, consenting Saudi adult participants (166 males and 70 females) were recruited and screened for the presence of the metabolic syndrome (MetS) using the modified National Cholesterol Education Program–Adult Treatment Panel III definition. Blood thiamine and its derivatives were quantified using HPLC.

Results: A total of 140 participants (53.9%) had MetS. The levels of thiamine and its derivatives of those with MetS were not significantly different from those without. However, hypertensive subjects had significantly higher urinary thiamine ($P = 0.03$) as well as significantly lower levels of thiamine diphosphate (TDP) ($P = 0.01$) and total thiamine ($P = 0.02$) than the normotensive subjects, even after adjusting for age and body mass index (BMI). Furthermore, age- and BMI-matched participants with elevated blood glucose levels had significantly lower levels of thiamine monophosphate ($P = 0.020$), TDP ($P < 0.001$) and total thiamine ($P < 0.001$) and significantly elevated levels of urinary thiamine ($P = 0.005$) compared to normoglycemic participants.

Conclusions: Low thiamine levels are associated with elevated blood glucose and hypertension in Saudi adults. Determination of thiamine status may be considered and corrected among patients with, or at high risk for, MetS, but the question whether thiamine deficiency correction translates to improved cardiometabolic status needs further longitudinal investigation.

Keywords: Arabs, Metabolic syndrome, Thiamine

Background

Thiamine (vitamin B1) is an essential water-soluble vitamin with major functions in carbohydrate metabolism. It is phosphorylated by thiamine kinase, and the major fraction of tissue thiamine is represented by thiamine diphosphate (TDP) [1]. This active form functions as a co-factor for both pyruvate (PDHC) and α -ketoglutarate dehydrogenase complexes (KDHCs). Studies have shown that decreased activity of PDHC, KDHC or succinate dehydrogenase secondary to thiamine deficiency results in

acetyl-coenzyme A and energy deficits in brain, muscle and other tissues [2-5].

The TDP concentration in erythrocytes has been observed to be a good indicator of body stores because it depletes at a rate similar to TDP levels found in major organs of the body [6]. Studies have suggested that erythrocyte TDP concentrations determined by both the apoenzyme recombination technique and high-performance liquid chromatography (HPLC) are more sensitive indices of thiamine status than measurement of erythrocyte transketolase activity [7,8]. The use of HPLC for the direct determination of TDP, however, has clear advantages in terms of sensitivity, specificity, precision and robustness [7-9].

HPLC methods for the determination of thiamine and its esters in blood have been documented [10]. We have

* Correspondence: aldaghri2011@gmail.com

¹Biomarkers Research Program, College of Science, King Saud University, Riyadh 11451, Kingdom of Saudi Arabia

²Prince Mutaib Chair for Biomarkers of Osteoporosis, King Saud University, Riyadh 11451, Kingdom of Saudi Arabia

Full list of author information is available at the end of the article

previously developed a step-gradient HPLC method to overcome the weaknesses of previously reported thiamine assays in terms of speed and accuracy for baseline separation of thiamine compounds using whole blood instead of washed erythrocytes and a simpler mobile phase [11]. In the present study, we aim to determine associations of thiamine and its derivatives with cardiometabolic parameters in a Saudi cohort with or without metabolic syndrome (MetS) using our method of thiamine quantification. The study will be one of the first to shed light on the association of thiamine with MetS parameters among the Saudi Arab ethnicity, in which MetS manifestations and other chronic noncommunicable diseases are highly prevalent [12,13].

Methods

Participants

Samples were taken from 236 randomly selected, healthy, consenting Saudi adult participants (166 males and 70 females) who were part of the Biomarkers Screening in

Riyadh (BSR), a capital-wide survey spanning all primary care centers in Riyadh initiated by the Biomarkers Research Program (BRP) of King Saud University (KSU) and the Ministry of Health. In brief, BSR was launched to identify and employ novel biomarkers of chronic noncommunicable diseases, including diabetes mellitus (DM), cardiovascular disease (CVD), hypertension and obesity, among consenting and randomly recruited Saudis. Ethical approval was obtained from the Ethics Committee of the College of Science Research Center of KSU, Riyadh, Saudi Arabia [12]. They were screened for the presence of MetS based on the definition of the modified National Cholesterol Education Program—Adult Treatment Panel III.

Reagents

A certified thiamine hydrochloride standard was obtained from Dr Ehrenstorfer GmbH (Augsburg, Germany). Thiamine monophosphate (TMP), TDP, potassium ferricyanide, trichloroacetic acid (TCA) and sodium hydroxide were obtained from Sigma Chemical Company (St Louis,

Table 1 General characteristics of participants with or without metabolic syndrome^a

Characteristics	Control	MetS	P-value
N	96	140	
Gender (M/F)	69/27	97/43	
Age (yr)	49.4 ± 16.5	53.7 ± 13.3	0.03
BMI (kg/m ²)	27.6 ± 6.0	31.3 ± 6.3	< 0.001
Waist circumference (cm)	81.5 ± 23.8	106.1 ± 12.5	< 0.001
Hip circumference (cm)	91.5 ± 26.8	112.1 ± 15.7	< 0.001
Sagittal abdominal diameter (cm)	22.3 ± 9.2	26.7 ± 7.1	< 0.001
Systolic blood pressure (mmHg)	124.3 ± 16.8	130.0 ± 14.3	0.01
Diastolic blood pressure (mmHg)	78.8 ± 10.5	81.0 ± 8.5	0.10
Triglycerides (mmol/L)	1.6 ± 0.13	1.9 ± 0.11	0.02
Total cholesterol (mmol/L)	5.0 ± 1.1	5.3 ± 1.2	0.12
LDL cholesterol (mmol/L)	3.5 ± 1.0	3.7 ± 1.0	0.18
HDL cholesterol (mmol/L)	0.70 ± 0.29	0.63 ± 0.24	0.07
Glucose (mmol/L)	7.4 ± 1.6	9.0 ± 1.5	0.01
Thiamine (ng/ml)	3.3 ± 0.13	3.2 ± 0.16	0.88
TMP (ng/ml)	2.1 ± 1.4	2.3 ± 1.5	0.19
TDP (ng/ml)	27.4 ± 1.8	22.3 ± 2.0	0.06
Total thiamine (ng/ml)	35.0 ± 1.8	31.3 ± 2.3	0.20
Thiamine (urine) (µg/ml)	938.0 ± 170.8	947.1 ± 207.4	0.94
Albumin (serum) (g/L)	47.8 ± 5.6	46.7 ± 6.4	0.31
Albumin (urine) (mg/L)	23.8 ± 4.1	16.7 ± 3.3	0.03
Serum creatinine (mmol/L)	78.6 ± 1.2	81.7 ± 1.5	0.52
Urinary creatinine (mmol/L)	11,966.8 ± 1,008.7	9,662.9 ± 1,393.6	0.07
Calcium (mmol/L)	2.6 ± 0.51	2.6 ± 0.50	0.78
Phosphorous (mmol/L)	1.3 ± 0.36	1.4 ± 0.43	0.25

Note: Data represent means ± standard deviation. Independent *t*-tests were done for two groups. *P* ≤ 0.05 was used as the significance level. ^aBMI body mass index, HDL high-density lipoprotein, LDL low-density lipoprotein, MetS metabolic syndrome, TDP thiamine diphosphate, TMP thiamine monophosphate.

MO, USA). HPLC-grade acetonitrile and methanol were purchased from VWR International (VWR International, Lutterworth, UK).

Urine sample preparation

Urine samples were extracted according to a previously described method [14]. In summary, samples were thawed in the dark at room temperature. After brief mixing, 0.1 ml of urine samples were transferred to a 2-ml amber glass tube and 0.9 ml of 0.01 M HCl was added, followed by vortex mixing. The resulting solution was then filtered through a Millipore polytetrafluoroethylene 0.45- μ m/25-mm filter (EMD Millipore, Billerica, MA, USA) into light-protected vials and placed in the autosampler tray of the HPLC for analysis.

Blood sample preparation

Fasting blood was drawn (about 10 ml), centrifuged and processed on the same day the participants submitted their consent forms. Both whole blood and serum were placed in plain polystyrene tubes. Collected fasting blood samples were delivered to BRP for storage at -20°C . On the day of analysis, blood samples were thawed at room temperature. An aliquot of 300 μ l of the sample was added to 50 μ l of TCA (TCA concentration = 50%). This aliquot was mixed using a vortex shaker for 1 min and centrifuged at 6,000 rpm/10 min. The resulting supernatant (250 μ l) was collected and transferred into clean 300 μ l microautosampler vials. A freshly prepared derivatizing reagent (0.2% potassium ferricyanide prepared in 15% sodium hydroxide) will subsequently be added (20 μ l) to the samples before injection.

Instrumental analysis

The BRP, College of Science, KSU, Riyadh, Saudi Arabia, has an HPLC system (Shimadzu Corp, Kyoto, Japan) that is fully equipped with a model series (LC-10ADVP pump, DGU-14A degasser, SIL-10ADVP autosampler, SPD-M20A UV/VIS detector, RF-10AXL fluorescence detector and SCL-10A system controller; Shimadzu Corp). System control and data analysis were carried out using LCsolution software (Shimadzu Corp). Separation of these analytes has been done on a Symmetry C18 column (5 μ m; 250 mm \times 4.6 mm; Waters Corp, Milford, MA, USA). The separation was carried out using a gradient elution procedure. Mobile phase A (30 mM sodium dihydrogen phosphate buffer adjusted to pH 4.5 and acetonitrile; 94:6 vol/vol ratio) and mobile phase B was 30 mmol/L sodium dihydrogen phosphate buffer (pH 4.5) and acetonitrile (70:30 vol/vol) ratios were linearly changed as follows: 0 to 3 min, 0%:30% mobile phase B; 3 to 5 min, 30%:90% mobile phase B; 5 to 9 min, 90% mobile phase B; 9 to 10 min, 90%:0% mobile phase B; and 10 to 20 min, 0.0% mobile phase B. The total run time was 20 min at a flow rate of 0.7 ml/min. The eluent was

monitored by a diode array detector and detection wavelength was set at 254 nm. Detection of thiamine was observed after precolumn derivatization at the excitation and emission wavelengths of 365 and 435 nm, respectively, with an injection volume of 50 μ l.

Chromatographic conditions, as well as analytical validation, which includes linearity and detection limit, recovery, precision and accuracy, have been described previously [11]. In brief, the standard calibration curves showed good linearity for thiamine at 0, 25, 50, 100, 150 and 200 ng/ml and 0, 10, 20, 40 and 80 ng/ml for both TMP and TDP, respectively. Correlation coefficient values are $r > 0.98$ for thiamine, $r > 0.97$ for TMP and $r > 0.99$ for TDP. Recovery percentages were $93.0\% \pm 4.0\%$, $97.0\% \pm 5.4\%$ and $91.0\% \pm 3.4\%$ for the three analytes, respectively.

Statistical analysis

Data were analyzed using the SPSS version 16.5 software (SPSS, Chicago, IL, USA). Continuous variables are presented as means \pm standard deviation, and variables exhibiting non-Gaussian distribution were normalized prior to analysis. Independent t -tests were used to compare groups between those with and without MetS. The same

Table 2 Age- and body mass index-adjusted comparisons between normotensive and hypertensive participants^a

Characteristics	Normotensive	Hypertensive	P-value
N	117	119	
Gender (M/F)	81/36	85/34	
Systolic blood pressure (mmHg)	114.76 \pm 8.85	137.67 \pm 11.5	< 0.001
Diastolic blood pressure (mmHg)	73.6 \pm 6.99	85.25 \pm 7.6	< 0.001
Triglycerides (mmol/L)	1.3 \pm 0.03	1.4 \pm 0.03	0.02
Total cholesterol (mmol/L)	5.0 \pm 1.0	5.4 \pm 1.2	0.12
LDL cholesterol (mmol/L)	3.5 \pm 0.10	3.7 \pm 0.10	0.20
HDL cholesterol (mmol/L)	0.62 \pm 0.03	0.70 \pm 0.03	0.08
Glucose (mmol/L)	8.7 \pm 0.41	9.9 \pm 0.41	0.04
Thiamine (ng/ml)	3.4 \pm 0.05	3.1 \pm 0.05	0.30
TMP (ng/ml)	2.2 \pm 0.04	2.3 \pm 0.04	0.51
TDP (ng/ml)	27.7 \pm 1.1	21.2 \pm 1.1	0.01
Total thiamine (ng/ml)	35.0 \pm 1.1	27.4 \pm 1.1	0.02
Thiamine (urine) (μ g/ml)	835.6 \pm 2.0	1,049.6 \pm 2.0	0.03
Albumin (serum) (g/L)	46.8 \pm 0.73	47.2 \pm 0.80	0.75
Albumin (urine) (mg/L)	17.3 \pm 1.1	21.0 \pm 1.2	0.30
Serum creatinine (mmol/L)	80.7 \pm 0.20	80.6 \pm 0.20	0.99
Urinary creatinine (mmol/L)	9,748.2 \pm 20.0	10,774.4 \pm 21.0	0.44
Calcium (mmol/L)	2.5 \pm 0.47	2.7 \pm 0.50	0.07
Phosphorous (mmol/L)	1.3 \pm 0.41	1.5 \pm 0.40	0.04

Note: Data represent means \pm standard deviation. Level of significance was set at $P \leq 0.05$. ^aHDL high-density lipoprotein, LDL low-density lipoprotein, TDP thiamine diphosphate, TMP thiamine monophosphate.

Table 3 Age- and body mass index-adjusted comparisons between normoglycemic and hyperglycemic participants^a

Characteristics	Normoglycemic	Hyperglycemic	P-value
N	65	171	
Gender (M/F)	34/31	135/36	
Glucose (mmol/L)	4.6 ± 1.3	9.7 ± 1.4	< 0.001
Triglycerides (mmol/L)	1.6 ± 0.06	1.9 ± 0.03	0.23
Total cholesterol (mmol/L)	5.2 ± 0.20	5.1 ± 0.10	0.94
LDL cholesterol (mmol/L)	3.6 ± 1.0	3.6 ± 1.0	0.91
HDL cholesterol (mmol/L)	0.76 ± 0.04	0.62 ± 0.03	0.005
Thiamine (ng/ml)	3.5 ± 0.06	3.1 ± 0.04	0.14
TMP (ng/ml)	2.5 ± 0.06	2.1 ± 0.04	0.020
TDP (ng/ml)	36.2 ± 1.1	21.4 ± 1.0	< 0.001
Total thiamine (ng/ml)	41.7 ± 1.1	27.7 ± 1.0	< 0.001
Thiamine (urine) (µg/ml)	685.1 ± 3.3	1040.7 ± 2.0	0.005
Albumin (serum) (g/L)	46.2 ± 4.6	47.3 ± 6.4	0.18
Albumin (urine) (mg/L)	18.3 ± 1.2	19.2 ± 1.1	0.93
Serum creatinine (mmol/L)	78.8 ± 1.2	81.3 ± 2.0	0.68
Urine creatinine (mmol/L)	10,140.5 ± 43.9	10,261.7 ± 25.6	0.93
Calcium (mmol/L)	2.5 ± 0.08	2.6 ± 0.05	0.68
Phosphorous (mmol/L)	1.5 ± 0.07	1.4 ± 0.04	0.21

Note: Data represent means ± standard deviation. Level of significance was set at $P \leq 0.05$. ^aHDL high-density lipoprotein, LDL low-density lipoprotein, TDP thiamine diphosphate, TMP thiamine monophosphate.

test was employed between normotensive versus hypertensive and normoglycemic versus hyperglycemic, with adjustments for age and body mass index (BMI). Regression analysis was done to determine associations of interest among variables. P -value significance was set at < 0.05 .

Results

Characteristics of participants with or without MetS

Table 1 shows the general characteristics and unadjusted comparisons of participants with or without MetS. As expected, those who harbor MetS had significantly higher anthropometric measures, including BMI ($P < 0.001$), waist circumference ($P < 0.001$) and hip circumference ($P < 0.001$), as well as sagittal abdominal diameter (SAD) ($P < 0.001$) and systolic blood pressure ($P = 0.011$) than those without MetS. Among the biochemical parameters, those with MetS also had significantly higher levels of fasting blood glucose ($P = 0.010$) and triglycerides ($P = 0.025$), as well as significantly lower urinary albumin levels ($P = 0.030$), than those without MetS. The rest of the parameters, including thiamine and its derivatives, were comparable.

Differences in thiamine levels based on cardiometabolic parameters

We then compared parameters based on the presence of cardiometabolic risk factors. Table 2 shows age- and BMI-adjusted differences in the metabolic parameters of participants with or without hypertension. The hypertensive individuals had significantly higher levels of fasting blood

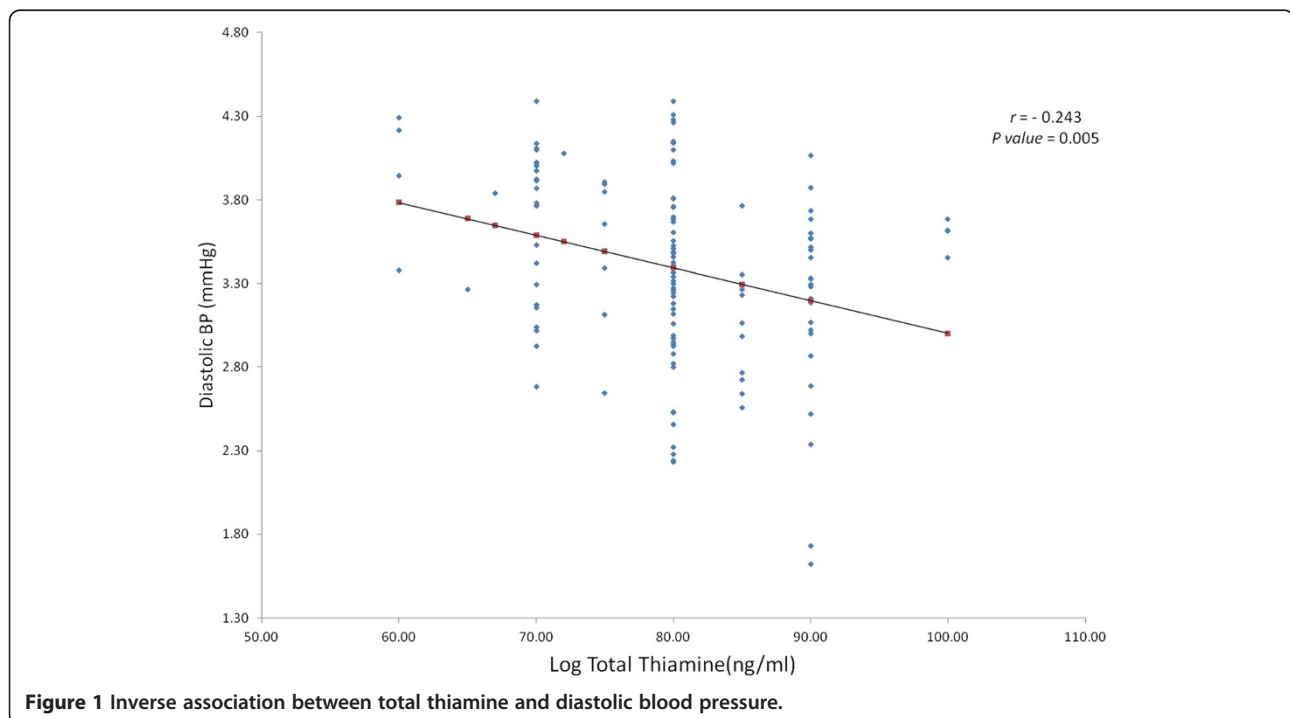


Figure 1 Inverse association between total thiamine and diastolic blood pressure.

glucose ($P = 0.04$), triglycerides ($P = 0.02$), phosphorous ($P = 0.04$) and urinary thiamine ($P = 0.03$), as well as significantly lower levels of TDP ($P = 0.01$) and total thiamine ($P = 0.02$), than the normotensive participants.

Table 3 shows age- and BMI-adjusted differences in the metabolic parameters of participants with or without elevated blood glucose. Those with elevated blood glucose had significantly lower levels of high-density lipoprotein (HDL) cholesterol ($P = 0.0055$), TMP ($P = 0.020$), TDP ($P < 0.001$) and total thiamine ($P < 0.001$), as well as significantly elevated levels of urinary thiamine ($P = 0.005$), than normoglycemic participants.

Comparisons of those with versus those without dyslipidemia, as well as those with elevated BMI, were also done, but they did not show significant differences in both serum and urinary thiamine levels. Figure 1 shows the modest but significant inverse association of serum total thiamine to diastolic blood pressure ($R = -0.24$, P value = 0.005). Total thiamine was also associated with age ($R = -0.16$, $P = 0.04$); glucose ($R = -0.32$, $P < 0.001$) and HDL cholesterol ($R = -0.17$, $P = 0.04$) (not shown in tables).

Discussion

The present cross-sectional study is the first to highlight the significant inverse associations of total thiamine to cardiometabolic parameters in a Saudi Arab cohort with or without the manifestations of MetS. MetS, being a complex disorder of interconnected risk factors, has consistently been tied to increased risk of atherosclerotic CD and diabetes mellitus type 2 (DMT2) [15]. Several recent studies done in Nigeria highlighted that water-soluble vitamins, including thiamine, and antioxidants are significantly lower in people with MetS [15,16]. Thiamine deficiency has been implicated in diabetic complications and other vascular diseases [17,18]. Its relation to blood pressure can be explained through its renal excretion. Being a water-soluble vitamin, it is normally salvaged by reabsorption from the glomerular filtrate [19], which is altered in the presence of an underlying renal pathology, a common cause of hypertension.

Thiamine deficiency has also been noted to be common among Saudi patients with DMT2 [11], and high-dose thiamine therapy has been shown to improve the condition of diabetic patients with renal complications, making it a promising adjuvant therapy for DMT2 patients with known thiamine deficiency [20]. The association of thiamine levels with glucose has been documented elsewhere, but not in select lipid parameters such as HDL cholesterol [21]. Animal studies, however, show correction of triglycerides and total and LDL cholesterol after high-dose thiamine supplementation, still with the exception of HDL cholesterol, but nevertheless suggesting possible cardioprotective effects, probably secondary to food intake suppression and hexosamine pathway signaling [22-24].

The authors acknowledge several limitations of this study. The results of the present study are at most suggestive, because the study design is cross-sectional. Furthermore, other confounders, such as physical activity and diet, were not taken into consideration, so we cannot rule out possible residual confounders with the analysis done. Nevertheless, this study is the first to document the association of circulating thiamine to cardiometabolic risk factors in an Arab cohort, suggesting that micronutrient deficiencies such as thiamine deficiency have cardiovascular implications if not corrected. Whether the present findings apply to Arab children also needs to be investigated because metabolic traits are highly heritable in this particular population [25]. Further studies that are interventional in design should be considered to validate findings.

Conclusions

Circulating thiamine and its derivatives are associated with blood glucose and blood pressure among Saudi Arabs with manifestations of MetS. Determination of thiamine status and correction thereof among patients with, or at high risk for, MetS may be considered. Longitudinal studies are needed to determine whether correction of thiamine deficiency can confer a favorable metabolic profile among those at risk for developing DMT2.

Competing interests

The authors declare no competing interests.

Authors' contributions

NMA and OSA conceptualized the study. KMA, MSA and SHA performed sample, as well as data collection, analysis and interpretation. SS wrote the manuscript. All authors approved the final version of the manuscript.

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Author details

¹Biomarkers Research Program, College of Science, King Saud University, Riyadh 11451, Kingdom of Saudi Arabia. ²Prince Mutaib Chair for Biomarkers of Osteoporosis, King Saud University, Riyadh 11451, Kingdom of Saudi Arabia. ³Center of Excellence in Biotechnology Research Center, King Saud University, Riyadh 11451, Kingdom of Saudi Arabia. ⁴Clinical Pharmacy Department, College of Pharmacy, King Saud University, Riyadh 11451, Kingdom of Saudi Arabia.

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References

1. Brown SB, Fitzsimons JD, Palace VP, Vandenbyllaardt L: Thiamine and early mortality syndrome in lake trout. *Am Fish Soc Symp* 1998, **21**:18-25.
2. Aikawa H, Watanabe IS, Furuse T, Iwasaki Y, Satoyoshi E, Sumi T: Low energy levels in thiamine-deficient encephalopathy. *J Neuropath Exp Ther* 1981, **43**:276-287.
3. Bubber P, Ke ZJ, Gibson GE: Tricarboxylic acid cycle enzymes following thiamine deficiency. *Neurochem Int* 2004, **45**:1021-1028.
4. Butterworth RF: Thiamine deficiency-related brain dysfunction in chronic liver failure. *Metab Brain Dis* 2009, **24**:189-196.
5. Langlais PJ: Pathogenesis of diencephalic lesions in an experimental model of Wernicke's encephalopathy. *Metab Brain Dis* 1995, **10**:31-44.

6. Brin M: Erythrocyte as biopsy tissue for functional evaluation of thiamin adequacy. *JAMA* 1964, **187**:762–766.
7. Baines DG: The evaluation of erythrocyte thiamin diphosphate as an indicator of thiamin status in man and its comparison with erythrocyte transketolase activity measurements. *Ann Clin Biochem* 1988, **25**:698–705.
8. Warnock LG, Prudhomme CR, Wagner C: The determination of thiamin pyrophosphate in blood and other tissues and its correlation with erythrocyte transketolase activity. *J Nutr* 1978, **108**:421–427.
9. Baines M: Improved HPLC determination of thiamin diphosphate in erythrocytes. *Clin Chim Acta* 1985, **153**:43–48.
10. The Diabetes Control and Complications Trial Research Group: The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. *N Engl J Med* 1993, **329**:977–986.
11. Al-Attas OS, Al-Daghri NM, Alfadda AA, Abd Al-Rahman SH, Sabico S: Blood thiamine and derivatives as measured by high-performance liquid chromatography: levels and associations in DM patients with varying degrees of microalbuminuria. *J Endocr Invest* 2012, **35**:951–956.
12. Al-Daghri NM, Al-Attas OS, Alokail MS, Alkharfy KM, Yousef M, Sabico SL, Chrousos GP: Diabetes mellitus type 2 and other chronic non-communicable diseases in the central region, Saudi Arabia (Riyadh cohort 2): a decade of an epidemic. *BMC Med* 2011, **9**:76.
13. Al-Daghri NM, Al-Attas OS, Alokail MS, Alkharfy KM, Sabico SL, Chrousos GP: Decreasing prevalence of the full metabolic syndrome but a persistently high prevalence of dyslipidemia among adult Arabs. *PLoS One* 2010, **5**:e12159.
14. Tabrizi AB: Development of a cloud point extraction-spectrofluorimetric method for trace copper(II) determination in water samples and parenteral solutions. *J Hazard Mater* 2007, **139**:260–264.
15. Kassi E, Pervanidou P, Kaltsas G, Chrousos G: Metabolic syndrome: definitions and controversies. *BMC Med* 2011, **9**:48.
16. Odum EP, Wakwe VC: Plasma concentrations of water soluble vitamins in metabolic syndrome patients. *Niger J Clin Pract* 2012, **15**:442–447.
17. Odum EP, Ejilemele AA, Wakwe VC: Antioxidant status of type 2 diabetic patients in Port Harcourt, Nigeria. *Niger J Clin Pract* 2012, **15**:55–58.
18. Page GL, Laight D, Cummings MH: Thiamine deficiency in diabetes mellitus and the impact of thiamine replacement on glucose metabolism and vascular disease. *Int J Clin Pract* 2011, **65**:684–690.
19. Karachalias N, Babael-Jadidi R, Rabbani N, Thornalley PJ: Increased protein damage in renal glomeruli, retina, nerve, plasma and urine and its prevention by thiamine and benfotiamine therapy in a rat model of diabetes. *Diabetologia* 2010, **53**:1506–1516.
20. Ashokkumar B, Vaziri ND, Said HM: Thiamin uptake by the human-derived renal epithelial (HEK-293) cells: cellular and molecular mechanisms. *Am J Physiol Renal Physiol* 2006, **291**:F796–F805.
21. Rabbani N, Alam SS, Riaz S, Larkin JR, Akhtar MW, Shafi T, Thornalley PJ: High-dose thiamine therapy for patients with type 2 diabetes and microalbuminuria: a randomized, double-blind placebo-controlled pilot study. *Diabetologia* 2009, **52**:208–212.
22. González-Ortiz M, Martínez-Abundis E, Robles-Cervantes JA, Ramírez-Ramírez V, Ramos-Zavala MG: Effect of thiamine administration on metabolic profile, cytokines and inflammatory markers in drug-naïve patients with type 2 diabetes. *Eur J Nutr* 2011, **50**:145–149.
23. Naveed AK, Qamar T, Ahmad I, Raheem A, Malik MM: Effect of thiamine on lipid profile on diabetic rats. *J Coll Physicians Surg Pak* 2009, **19**:165–168.
24. Babael-Jadidi R, Karachalias N, Kupich C, Ahmed N, Thornalley PJ: High-dose thiamine therapy counters dyslipidemia in streptozotocin-induced diabetic rats. *Diabetologia* 2004, **47**:2235–2246.
25. Al-Daghri NM, Al-Attas OS, Alokail MS, Alkharfy KM, Yakout SM, Sabico SB, Gibson GC, Chrousos GP, Kumar S: Parent-offspring transmission of adipocytokine levels and their association with metabolic traits. *PLoS One* 2011, **6**:e18182.

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